

101.213.24 – including *Taq* pol., IFU-01.
101.213.24u – without *Taq* pol., IFU-02.

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“Instructions for Use” (IFU)

Lot No.: **6G3**

Lot-specific information
Olerup SSP[®] DQB1*02

Product number:	101.213-24 – including <i>Taq</i> polymerase 101.213-24u – without <i>Taq</i> polymerase
Lot number:	6G3
Expiry date:	2021-02-01
Number of tests:	24
Number of wells per test:	31+1
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

This Product Description is only valid for Lot No. 6G3.

Complete product documentation consists of generic Instructions for Use (IFU),
lot specific Product Insert, Worksheet and Certificate

**CHANGES COMPARED TO THE PREVIOUS OLERUP SSP[®]
DQB1*02 LOT (8E9)**

The DQB1*02 kit is updated to enable separation of:

- Null and Alternatively expressed alleles
- The product documentation has been updated for new alleles of IMGT 3.32.0

The format of the Worksheet has been changed.

The DQB1*02 primer set, specificity and interpretation tables have been updated with the DQB1 alleles described since the previous *Olerup SSP[®] DQB1*02 lot (Lot No. 8E9)* was made.

Primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
5	Added	-	5'-primer added for the DQB1*02:96N allele.
11	Exchanged	Exchanged	5'-primer and 3'-primer exchanged for decreased tendency of primer oligomer formation.
28	Added	-	5'-primer added for the DQB1*02:96N allele.

101.213.24 – including *Taq* pol., IFU-01.
101.213.24u – without *Taq* pol., IFU-02.

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“Instructions for Use” (IFU)

Lot No.: 6G3

Lot-specific information

Change in revision R01 compared to R00:

1. Primer mix 8 does not amplify the DQB1*02:10 and 02:81 and the DQB1*03:49 and 05:152 alleles. Thus, this lot of the DQB1*02 kit cannot distinguish the DQB1*02:10 and the DQB1*02:02:01:01-02:02:05, 02:65, 02:95, 02:97 alleles or the DQB1*02:81 and the DQB1*02:01:01-02:01:24, 02:43-02:45, 02:47, 02:49, 02:52, 02:55-02:57, 02:60-02:61, 02:63, 02:66, 02:68-02:71, 02:73-02:76, 02:78, 02:82-02:83, 02:85-02:88, 02:90-02:94, 02:98-02:106 alleles. This has been corrected in the Interpretation and Specificity Tables.

Changes in revision R02 compared to R01:

1. Primer mix 7 does not amplify the DQB1*02:06, 02:48, 03:24, and 03:79 alleles.
 The corrections above have been implemented in the Specificity and Interpretation tables.

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“Instructions for Use” (IFU)

Lot No.: **6G3**

Lot-specific information

Well **32** contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup* SSP[®] HLA Class I, DRB, DQB1, DPB1 and DQA1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

HLA-specific PCR product sizes range from 75 to 200 base pairs.
The PCR product generated by the positive control primer pair is 430 base pairs.

Length of PCR product	105	200	105	80	75	80	85
5'-primer¹	164	340	440	45	45	43	36
	5'-CAC ^{3'}	5'-Agg ^{3'}	5'-TTA ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}	5'-TAC ^{3'}
							36
							5'-TAT ^{3'}
3'-primer²	231	2nd I	507	59	58	57	47
	5'-TgC ^{3'}	5'-AAA ^{3'}	5'-TTg ^{3'}	5'-CTC ^{3'}	5'-ggC ^{3'}	5'-CTC ^{3'}	5'-ACA ^{3'}
							48
							5'-gCA ^{3'}
							48
							5'-gCC ^{3'}
							52
							5'-TgT ^{3'}
A*	+	+	+				
B*	+	+	+				
C*	+	+	+				
DRB1				+	+		
DRB3				+	+		
DRB5				+			
DQB1					+		
DPB1						+	
DQA1							+

¹The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

²The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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Lot No.: **6G3**

Lot-specific information

PRODUCT DESCRIPTION

DQB1*02 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the DQB1*02:01 to DQB1*02:106 alleles.

Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.

PLATE LAYOUT

Each test consists of 32 PCR reactions in a 32 well PCR plate.

1	2	3	4	5	6	7	8
9	10	11	12	13	14	15	16
17	18	19	20	21	22	23	24
25	26	27	28	29	30	31	NC

The 32 well PCR plate is marked with 'DQB1*02' in silver/gray ink.

Well No. 1 is marked with the Lot No. '6G3'.

Wells 1 to 31 – DQB1*02 high resolution primers.

Well 32 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded. The PCR plates are heat-sealed with a PCR-compatible foil.

Please note: When removing each 32 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Due to the sharing of sequence motifs between DQB1 alleles, non-DQB1*02 alleles will be amplified by some primer mixes. For further details see Specificity Table.

UNIQUELY IDENTIFIED ALLELES

All the DQB1*02 alleles, i.e. **DQB1*02:01 to DQB1*02:106**, recognized by the HLA Nomenclature Committee in April 2018^{1,2} will be amplified by the primers in the DQB1*02 subtyping kit^{1,2,3}.

The DQB1*02 kit enables separation of the confirmed DQB1*02 alleles as listed in the IMGT/HLA database 3.26.0. An HLA allele is listed as confirmed by IMGT/HLA if it has been sequenced by more than a single laboratory or from multiple sources. Current allele confirmation status for DQB1*02 alleles is listed below.

101.213.24 – including *Taq* pol., IFU-01.
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Lot No.: **6G3**

Lot-specific information

The DQB1*02 kit also enables identification of polymorphisms in exons outside of the region encoding the peptide binding domain and of null and alternatively expressed alleles.

The following DQB1*02 alleles can be distinguished by the different sizes of the specific PCR product:

Alleles	Primer mix	Alleles	Primer mix
DQB1*02:07:01-02:07:02, 02:16	9	DQB1*02:20N, 02:22	16
DQB1*02:09, 02:24	11	DQB1*02:21, 02:35	15
DQB1*02:15, 02:29	18	DQB1*02:27, 02:28	22
DQB1*02:18N, 02:34	14	DQB1*02:41, 02:53Q	23

¹HLA-DQB1 alleles listed on the IMGT/HLA web page 2018-April-16, release 3.32.0, www.ebi.ac.uk/imgt/hla.

²Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page <http://hla.alleles.org/alleles/deleted.html>.

³This lot of the DQB1*02 kit cannot distinguish the DQB1*02:10 and the DQB1*02:02:01:01-02:02:05, 02:65, 02:95, 02:97 alleles or the DQB1*02:81 and the DQB1*02:01:01-02:01:24, 02:43-02:45, 02:47, 02:49, 02:52, 02:55-02:57, 02:60-02:61, 02:63, 02:66, 02:68-02:71, 02:73-02:76, 02:78, 02:82-02:83, 02:85-02:88, 02:90-02:94, 02:98-02:106 alleles.

RESOLUTION IN HOMO- AND HETEROZYGOTES

Results file with resolution in DQB1*02 homo- and heterozygotes is available upon request.

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Lot No.: **6G3**

Lot-specific information

ALLELE CONFIRMATION STATUS

Allele	Status ¹	Allele	Status ¹	Allele	Status ¹
DQB1*02:01:01	Confirmed	DQB1*02:14:01	Confirmed	DQB1*02:53Q	Confirmed
DQB1*02:01:02	Unconfirmed	DQB1*02:14:02	Unconfirmed	DQB1*02:54	Confirmed
DQB1*02:01:03	Unconfirmed	DQB1*02:15	Unconfirmed	DQB1*02:55	Unconfirmed
DQB1*02:01:04	Confirmed	DQB1*02:16	Confirmed	DQB1*02:56	Unconfirmed
DQB1*02:01:05	Confirmed	DQB1*02:17	Confirmed	DQB1*02:57	Unconfirmed
DQB1*02:01:06	Unconfirmed	DQB1*02:18N	Confirmed	DQB1*02:58N	Unconfirmed
DQB1*02:01:07	Confirmed	DQB1*02:19	Confirmed	DQB1*02:59	Unconfirmed
DQB1*02:01:08	Confirmed	DQB1*02:20N	Confirmed	DQB1*02:60	Unconfirmed
DQB1*02:01:09	Unconfirmed	DQB1*02:21	Unconfirmed	DQB1*02:61	Unconfirmed
DQB1*02:01:10	Confirmed	DQB1*02:22	Unconfirmed	DQB1*02:62	Confirmed
DQB1*02:01:11	Confirmed	DQB1*02:23	Confirmed	DQB1*02:63	Unconfirmed
DQB1*02:01:12	Unconfirmed	DQB1*02:24	Unconfirmed	DQB1*02:64	Unconfirmed
DQB1*02:01:13	Unconfirmed	DQB1*02:25	Unconfirmed	DQB1*02:65	Unconfirmed
DQB1*02:01:14	Confirmed	DQB1*02:26	Confirmed	DQB1*02:66	Unconfirmed
DQB1*02:01:15	Unconfirmed	DQB1*02:27	Confirmed	DQB1*02:67N	Unconfirmed
DQB1*02:01:16	Unconfirmed	DQB1*02:28	Confirmed	DQB1*02:68	Unconfirmed
DQB1*02:01:17	Unconfirmed	DQB1*02:29	Confirmed	DQB1*02:69	Unconfirmed
DQB1*02:01:18	Unconfirmed	DQB1*02:30	Confirmed	DQB1*02:70	Unconfirmed
DQB1*02:01:19	Confirmed	DQB1*02:31	Unconfirmed	DQB1*02:71	Unconfirmed
DQB1*02:01:20	Confirmed	DQB1*02:32	Unconfirmed	DQB1*02:72	Unconfirmed
DQB1*02:01:21	Unconfirmed	DQB1*02:33	Confirmed	DQB1*02:73	Unconfirmed
DQB1*02:01:22	Unconfirmed	DQB1*02:34	Unconfirmed	DQB1*02:74	Unconfirmed
DQB1*02:01:23	Confirmed	DQB1*02:35	Unconfirmed	DQB1*02:75	Unconfirmed
DQB1*02:01:24	Unconfirmed	DQB1*02:36	Unconfirmed	DQB1*02:76	Unconfirmed
DQB1*02:02:01:01	Confirmed	DQB1*02:37	Unconfirmed	DQB1*02:77	Unconfirmed
DQB1*02:02:01:02	Unconfirmed	DQB1*02:38	Unconfirmed	DQB1*02:78	Unconfirmed
DQB1*02:02:02	Unconfirmed	DQB1*02:39	Unconfirmed		
DQB1*02:02:03	Unconfirmed	DQB1*02:40	Unconfirmed		
DQB1*02:03	Confirmed	DQB1*02:41	Confirmed		
DQB1*02:04	Unconfirmed	DQB1*02:42	Confirmed		
DQB1*02:05	Confirmed	DQB1*02:43	Unconfirmed		
DQB1*02:06	Unconfirmed	DQB1*02:44	Unconfirmed		
DQB1*02:07:01	Confirmed	DQB1*02:45	Unconfirmed		
DQB1*02:07:02	Unconfirmed	DQB1*02:46	Confirmed		
DQB1*02:08	Unconfirmed	DQB1*02:47	Unconfirmed		
DQB1*02:09	Unconfirmed	DQB1*02:48	Unconfirmed		
DQB1*02:10	Unconfirmed	DQB1*02:49	Unconfirmed		
DQB1*02:11	Unconfirmed	DQB1*02:50	Confirmed		
DQB1*02:12	Confirmed	DQB1*02:51	Confirmed		
DQB1*02:13	Unconfirmed	DQB1*02:52	Unconfirmed		

¹Allele status “confirmed” or “unconfirmed” as listed on the IMGT/HLA web page 2016-October-14, release 3.26.0, www.ebi.ac.uk/imgt/hla.

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“Instructions for Use” (IFU)

Lot No.: **6G3**

Lot-specific information

SPECIFICITY TABLE

DQB1*02 SSP subtyping

Specificities and sizes of the PCR products of the 31+1 primer mixes used for DQB1*02 SSP subtyping

Primer Mix	Size of spec. PCR product ¹	Size of control band ²	Amplified DQB1*02 alleles ³	Other amplified DQB1 alleles
1⁴	120 bp	515 bp	*02:01:01-02:02:05, 02:04-02:16, 02:18N-02:24, 02:26-02:39, 02:41-02:71, 02:73-02:76, 02:78-02:106	
2^{4,5}	85 bp	430 bp	*02:03, 02:77	
3⁵	145 bp	515 bp	*02:01:01-02:01:24, 02:04-02:05, 02:07:01-02:09, 02:13-02:25, 02:27-02:47, 02:49, 02:51-02:61, 02:63, 02:66-02:79, 02:81-02:83, 02:85-02:88, 02:90-02:94, 02:96N, 02:98-02:106	*03:01:01:01-03:01:01:12, 03:01:01:14-03:23:02, 03:25:01-03:78, 03:80-03:96, 03:98-03:163, 03:166-03:278, 03:280, 04:01:01:01-04:02:01:01, 04:02:01:04-04:42, 05:01:01:01-05:02:10, 05:02:12-05:13, 05:15-05:83, 05:85-05:160, 06:01:01:01-06:37, 06:39-06:85, 06:87-06:101, 06:105-06:217, 06:219, 06:221-06:226, 06:228-06:252N
4	140 bp	430 bp	*02:02:01:01-02:03, 02:06, 02:10-02:12, 02:26, 02:50, 02:62, 02:64-02:65, 02:95, 02:97	
5⁴	95 bp 140 bp 195 bp	430 bp	*02:23 *02:04, 02:37 *02:96N	
6	210 bp	430 bp	*02:05	
7	165 bp	430 bp	*02:19	
8	160 bp	430 bp		
9⁴	95 bp 195 bp	430 bp	*02:16 *02:07:01-02:07:02	
10	180 bp 230 bp	430 bp	*02:08 *02:12	
11⁴	95 bp 170 bp	430 bp	*02:24 *02:09	
12⁴	120 bp 160 bp	430 bp	*02:11, 02:25 *02:13, 02:30	
13⁴	120 bp 215 bp	430 bp	*02:17, 02:32 *02:38	
14	145 bp 220 bp	430 bp	*02:18N, 02:54 *02:34, 02:38	
15⁴	100 bp	515 bp	*02:21, 02:39	

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“Instructions for Use” (IFU)

Lot No.: **6G3**

Lot-specific information

16⁴	205 bp		*02:35	
	100 bp	430 bp	*02:22, 02:39	
	150 bp		*02:54	
	230 bp		*02:20N	
17	140 bp	430 bp	*02:26, 02:37, 02:48	*03:24, 03:79
18⁴	90 bp	430 bp	*02:15	
	185 bp		*02:29	
19	165 bp	430 bp	*02:33, 02:36	
20	165 bp	430 bp	*02:14:01-02:14:02, 02:30, 02:36	
21⁴	100 bp	430 bp	*02:23, 02:31	
	130 bp		*02:40	
22^{4,5}	90 bp	430 bp	*02:27	
	170 bp		*02:28, 02:59	*03:241, 06:44, 06:47
23⁴	100 bp	430 bp	*02:53Q	
	255 bp		*02:41	
24	160 bp	430 bp	*02:51	
	200 bp		*02:42	
25	160 bp	430 bp	*02:51	
	220 bp		*02:46	
26	185 bp	430 bp	*02:50	
27	155 bp	430 bp	*02:62	
	210 bp		*02:72	
28	150 bp	430 bp	*02:64, 02:79	*03:21, 05:35, 06:243
	195 bp		*02:96N	
29⁴	65 bp	430 bp	*02:67N	
30⁴	110 bp	430 bp	*02:58N	
31⁴	75 bp	430 bp	*02:59, 02:80	*03:241, 06:44, 06:47
32⁶	-	-	Negative Control	

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DQB1*02 SSP typings.

When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

³For several DQB1 alleles 1st and/or 3rd exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched

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“Instructions for Use” (IFU)

Lot No.: 6G3

Lot-specific information

with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

⁴HLA-specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

⁵Primer mixes 2, 3 and 22 may have tendencies of unspecific amplifications.

⁶Primer mix 32 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by the control primer pairs matching the human growth hormone gene. HLA-specific PCR product sizes range from 75 to 200 base pairs and the PCR product generated by the HGH positive control primer pair is 430 base pairs.

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“Instructions for Use” (IFU)

Lot No.: **6G3**

Lot-specific information

PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11	12
Length of spec.	120	85	145	140	95	210	165	160	95	180	95	120
PCR product					140				195	230	170	160
					195							
Length of int.	515	430	515	430	430	430	430	430	430	430	430	430
pos. control ¹												
5'-primer(s) ²	30(185) 5'-AAG 3'	57(266) 5'-TgA 3'	135(500) 5'-TgA 3'	102(400) 5'-TCT 3'	26(173) 5'-TTT 3'	21(159) 5'-ACA 3'	30(185) 5'-AAG 3'	102(400) 5'-TCT 3'	30(185) 5'-AAG 3'	30(185) 5'-AAG 3'	36(204) 5'-gAT 3'	16(142) 5'-gCA 3'
					39(212) 5'-gCT 3'		102(400) 5'-TCT 3'				135(500) 5'-TgA 3'	17(147) 5'-TTA 3'
					118(449) 5'-CTA 3'							30(184) 5'-gAg 3'
					136(502) 5'-ACg 3'							33(193) 5'-ATg 3'
3'-primer(s) ³	57(266) 5'-CAg 3'	71(309) 5'-CgT 3'	170(604) 5'-gAC 3'	135(500) 5'-ggC 3'	57(266) 5'-Cgg 3'	77(326) 5'-CCg 3'	72(311) 5'-CCg 3'	142(520) 5'-AAT 3'	49(241) 5'-CgT 3'	77(326) 5'-CCT 3'	55(260) 5'-gCA 3'	57(266) 5'-Cgg 3'
	57(266) 5'-Cgg 3'				170(604) 5'-gAC 3'		170(604) 5'-gAT 3'		81(338) 5'-Tgg 3'	92(372) 5'-CgA 3'	179(631) 5'-gCg 3'	
	58(270) 5'-TCA 3'								81(338) 5'-TCg 3'			
Well No.	1	2	3	4	5	6	7	8	9	10	11	12

Well No.	13	14	15	16	17	18	19	20	21	22	23	24
Length of spec.	120	145	100	100	140	90	165	165	100	90	100	160
PCR product	215	220	205	150		185			130	170	255	200
				230								
Length of int.	430	430	515	430	430	430	430	430	430	430	430	430
pos. control ¹												
5'-primer(s) ²	30(185) 5'-AAG 3'	30(185) 5'-AAG 3'	30(185) 5'-AAG 3'	30(185) 5'-AAG 3'	23(163) 5'-AgT 3'	9(122) 5'-gTg 3'	13(134) 5'-ggA 3'	13(134) 5'-ggA 3'	28(178) 5'-TgC 3'	13(134) 5'-ggT 3'	14(138) 5'-ATA 3'	30(185) 5'-AAG 3'
					26(173) 5'-TTT 3'	41(217) 5'-TCC 3'	20(154) 5'-ACT 3'	17(147) 5'-TTA 3'	38(209) 5'-CgC 3'	41(217) 5'-TCT 3'	66(292) 5'-..g 3'	
					135(500) 5'-TgA 3'			20(154) 5'-ACC 3'	39(212) 5'-gCT 3'	135(500) 5'-TgA 3'		
								20(154) 5'-ACA 3'				
3'-primer(s) ³	57(265) 5'-ggT 3'	61(279) 5'-TTT 3'	47(236) 5'-ggT 3'	47(236) 5'-ggT 3'	57(266) 5'-Cgg 3'	57(266) 5'-Cgg 3'	57(266) 5'-Cgg 3'	57(266) 5'-Cgg 3'	57(266) 5'-Cgg 3'	57(266) 5'-Cgg 3'	86(353) 5'-gCT 3'	69(303) 5'-CTg 3'
	58(268) 5'-ggT 3'	66(294) 5'-ACT 3'	51(248) 5'-gCC 3'	51(248) 5'-gCA 3'	170(604) 5'-gAT 3'					174(618) 5'-ACT 3'		83(343) 5'-gTg 3'
	88(359) 5'-TgT 3'	88(359) 5'-TgT 3'	84(348) 5'-AAA 3'	66(294) 5'-ACT 3'								
		93(374) 5'-gCg 3'		94(376) 5'-TCA 3'								
Well No.	13	14	15	16	17	18	19	20	21	22	23	24

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101.213.24u – without *Taq* pol., IFU-02.

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“Instructions for Use” (IFU)

Lot No.: **6G3**

Lot-specific information

Well No.	25	26	27	28	29	30	31
Length of spec.	160	185	155	150	65	110	75
PCR product	220		210	195			
Length of int.	430	430	430	430	430	430	430
pos. control ¹							
5'-primer(s) ²	30(185) 5'-AAg 3'	38(208) 5'-TCA 3'	30(185) 5'-AAA 3'	118(449) 5'-CTA 3'	9(122) 5'-gTA 3'	35(199) 5'-gAT 3'	174(618) 5'-CAA 3'
			48(239) 5'-CCA 3'	133(493) 5'-TTT 3'			
3'-primer(s) ³	69(303) 5'-CTg 3'	86(353) 5'-gCT 3'	86(353) 5'-gCT 3'	170(604) 5'-gAC 3'	16(144) 5'-AAC 3'	57(266) 5'-Cgg 3'	185(650) 5'-Cgg 3'
	89(362) 5'-TCT 3'						
Well No.	25	26	27	28	29	30	31

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

²The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

³The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

101.213.24 – including *Taq* pol., IFU-01.
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Lot No.: 6G3

Lot-specific information

CELL LINE VALIDATION SHEET																				
DQB1*02 SSP subtyping kit ²																				
					Well															
				Production No.	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
					201558101	201558102	201443803	2014443804	201894505	2014443806	201443807	201777208	2014443809	2014443810	201894511	201777212	2014443813	201558114	201777215	201558116
	IHWC cell line ¹		DQB1																	
1	9001	SA	*05:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
2	9280	LK707	*06:01	*02:02	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-
3	9011	E4181324	*06:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
4	9275	GU373	*02:01		+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
5	9009	KAS011	*05:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
6	9353	SM	*03:02	*06:01	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
7	9020	QBL	*02:01		+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
8	9025	DEU	*03:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
9	9026	YAR	*03:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
10	9107	LKT3	*04:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
11	9051	PITOUT	*02:02		+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
12	9052	DBB	*03:03		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
13	9004	JESTHOM	*05:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
14	9071	OLGA	*04:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
15	9075	DKB	*03:03		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
16	9037	SWEIG007	*03:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
17	9282	CTM3953540	*02:01	*06:03	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
18	9257	32367	*06:02	*02:02	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-
19	9038	BM16	*03:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
20	9059	SLE005	*06:04		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
21	9064	AMALA	*03:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
22	9056	KOSE	*05:03	*06:04	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
23	9124	IHL	*05:03	*06:01	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
24	9035	JBUSH	*03:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
25	9049	IBW9	*02:02		+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
26	9285	WT49	*02:01		+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
27	9191	CH1007	*04:01	*05:01	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
28	9320	BEL5GB	*02:02	*03:01	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-
29	9050	MOU	*02:02		+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
30	9021	RSH	*04:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
31	9019	DUCAF	*02:01		+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
32	9297	HAG	*03:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
33	9098	MT14B	*03:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
34	9104	DHIF	*03:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
35	9302	SSTO	*03:05		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
36	9024	KT17	*03:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
37	9065	HHKB	*06:03		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
38	9099	LZL	*03:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
39	9315	CML	*02:01	*03:01	+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
40	9134	WHONP199	*02:02	*03:03	+	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-
41	9055	H0301	*06:09		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
42	9066	TAB089	*06:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
43	9076	T7526	*03:03		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
44	9057	TEM	*05:03		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
45	9239	SHJO	*02:02		+	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
46	9013	SCHU	*06:02		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
47	9045	TUBO	*03:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
48	9303	TER-ND	*05:01		-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-

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Lot-specific information

[illegible]

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Lot-specific information

¹The provided cell line HLA specificities are retrieved from the <http://www.ihwg.org/hla> web site. The specificity of an individual cell line may thus be subject to change.

²The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 5 to 31 were available. The specificities of the primers in primer solutions 6, 11, 15 to 17 and 22 were tested by separately adding one 5'-primer, respectively one 3'-primer. In primer solutions 5, 12, 18 to 21, 23, 26 to 28, 30 and 31 it was only possible to test the 3'-primers, the 5'-primers were not possible to test. In primer solutions 7 to 10, 13, 14, 24, 25 and 29 it was only possible to test the 5'-primers, the 3'-primers were not possible to test.

In primer solutions 11, 17 and 22 one or two 5'-primers were not possible to test, and in primer solutions 1, 11, 15 to 17 and 22 one, two or three 3'-primers were not possible to test.

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Lot No.: **6G3**

Lot-specific information

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